<u>A proposed molecular mechanism for pathogenesis of</u> <u>severe RNA-viral pulmonary infections</u>

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SRSF1 and RNPS1 Information Models and Binding Sites in Host and Viral Genomes

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Factor		<u>SRSF1 [Rep1]^{1,2}</u>		<u>SRSF1 [Rep1] /</u> <u>RNPS1 Model</u> <u>Comparison</u>		RNPS1 ¹		SRSF1 [Rep2] / RNPS1 Model Comparison		<u>SRSF1 [Rep2]^{1,2}</u>	
Sequence Logo				-				-			
R _{sequence} (bits)		6.7 ± 2.1		-		7.8 ± 1.9		-		6.4 ± 2.1	
Motif Similarity (E-value) ³		-		5.0e-09		-		1.1e-09		-	
No. of Expressed Binding Sites (A549; ≥ 0 bits) ⁴		1.3e08		5.4e07 (57%) ⁷		9.3e07		6.4e07 (69%)		1.5e08	
No. of Expressed Binding Sites (Pneumocytes; ≥0 bits) ⁵		6.8e07		2.9e07 (58%)		5.0e07		3.4e07 (69%)		7.9e07	
No. of Sites (SARS-CoV- 2; + - strand)	≥ 0 bits	860	732	435 (72%)	305 (65%)	608	466	363 (60%)	273 (59%)	810	772
	$\geq 1/2$ Rseq	311	232	131 (51%)	86 (44%)	256	196	155 (61%)	115 (59%)	376	358
	≥ Rseq	31	42	16 (46%)	10 (40%)	35	25	35 (100%)	25 (100%)	60	33
No. of Sites (Influenza A; + - strand) ⁶	≥ 0 bits	697	339	289 (61%)	118 (63%)	475	188	268 (56%)	129 (69%)	616	388
	≥ ¹/ ₂ Rseq	263	118	122 (49%)	47 (55%)	248	85	162 (65%)	65 (76%)	373	188
	≥ Rseq	50	23	24 (53%)	12 (75%)	45	16	45 (100%)	16 (100%)	84	35

¹ RNPS1 model derived from publicly available iCLIP data (E-MTAB-4215; ArrayExpress), while SRSF1 models were derived from eCLIP data (ENCSR456FVU; ENCODE Data Coordination Center); ² SRSF1 [Rep1] and [Rep2] were derived from eCLIP dataset replicate 1 [50,000 peaks] and replicate 2 [5,000 peaks], respectively; ³ RNA binding motifs were compared using STAMP (34) using the Pearson Correlation Coefficient distance metric (74); ⁴ A549 cell line expression from GSE141171 dataset; ⁵ Primary type II pneumocyte expression from GSE86618 dataset; ⁶ Influenza A virus H3N2 strain (Ontario/104-25/2012). ⁷ RNPS1 sites used as denominator for all percentages.

Distribution of SRSF1 and RNPS1 Binding Sites Across SARS-CoV-2



Positive Strand Binding Sites

Negative Strand Binding Sites

R_i of SRSF1 and RNPS1 Binding Sites in the SARS-CoV-2 and Human Genome



The SARS-CoV-2 Viral Genome

Transcribed Regions of the Human Genome

Expression of Rad-signature Genes in Influenza-infected and Radiation-Exposed Leukocytes



Inhibition of Host SRSF1 Binding by Viral Genome Replication



Summary

- The proposed mechanism of RNA viral infection-induced apoptosis is supported by bioinformatic analysis of RNA binding events
- Host RNA binding protein (RBP) binding sites are frequent and conserved among different strains of RNA viral genomes
- DNA repair and apoptotic proteins (such as DDB2, PCNA, PRKCH and GTF3A) were differentially expressed in influenza A and dengue-infected cells
- Information-dense binding site clustering of strong RBP binding sites coincides with the distribution of RNA-DNA hybridization sites across the genome
- Most binding sites and information-dense clusters do not overlap DRIP- and DRIPc-seq intervals, which are regions in the human genome with evidence of Rloop formation
- SRSF1 and RNPS1 have similar binding motifs which explains why DNA damage from SRSF1 depletion is complimented by RNPS1 overexpression

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RESEARCH ARTICLE

A proposed molecular mechanism for pathogenesis of severe RNA-viral pulmonary infections

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Abstract

Background: Certain riboviruses can cause severe pulmonary complications leading to death in some infected patients. We propose that DNA damage induced-apoptosis accelerates viral release, triggered by depletion of host RNA binding proteins (RBPs) from nuclear RNA bound to replicating viral sequences.

Methods: Information theory-based analysis of interactions between RBPs and individual sequences in the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), influenza A (H3N1), HIV-1, and dengue viral genomes identifies strong RBP binding sites in viral genomes. Replication and expression of viral sequences are expected to increasingly sequester RBPs - SRSF1 and RNPS1. Ordinarily, RBPs bound to nascent host transcripts prevents their annealing to complementary DNA. Their depletion induces destabilizing R-loops. Chromosomal breakage occurs when an excess of unresolved R-loops collide with incoming replication forks, overwhelming the DNA repair machinery. We estimated stoichiometry of inhibition of RBPs in host nuclear RNA by counting competing binding sites in replicating viral genomes and host RNA.

Results: Host RBP binding sites are frequent and conserved among different strains of RNA viral genomes. Similar binding motifs of SRSF1 and RNPS1 explain why DNA damage resulting from SRSF1 depletion is complimented by expression of RNPS1. Clustering of strong RBP binding sites coincides with the distribution of RNA-DNA hybridization sites across the genome. SARS-CoV-2 replication is estimated to require 32.5-41.8 hours to effectively compete for binding of an equal proportion of SRSF1 binding sites in host encoded nuclear RNAs. Significant changes in expression of transcripts encoding DNA repair and apoptotic proteins were found in an analysis of influenza A and Dengue-infected cells in some individuals.

Conclusions: R-loop-induced apoptosis indirectly resulting from viral replication could release significant quantities of membrane-associated virions into neighboring alveoli. These could infect adjacent pneumocytes and other tissues, rapidly compromising lung function, multiorgan system failure and the other described symptoms.

Reviewer Status AWAITING PEER REVIEW

article can be found at the end of the article.

Keywords

SARS-CoV-2, Influenza A, HIV-1, Dengue Virus, Apoptosis, R-loop, DNA damage, RNA binding protein

Alternative Versions of Slides

Distribution of SRSF1 and RNPS1 Binding Sites Across Influenza A



